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Title:

NOVEL HIV-SPECIFIC SYNTHETIC OLIGONUCLEOTIDES
AND METHODS OF THEIR USE

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NOVEL HIV-SPECIFIC SYNTHETIC OLIGONUCLEOTIDES
AND METHODS OF THEIR USE

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5 BACKGROUND OF THE INVENTION

 This invention relates to the treatment of
HIV infection. More particularly, this invention
relates to synthetic modified antisense
10 oligonucleotides and pharmaceutical compositions
containing such oligonucleotides and to methods of
inhibiting HIV replication and treating HIV
infection using such oligonucleotides.

15 Human immunodeficiency virus types 1 and 2
(HIV-1, HIV-2), formerly called human T-cell
leukemia lymphotropic virus-type III (HTLV-III),
are believed to be the etiological agents of
acquired immune deficiency syndrome (AIDS). HIV
20 is part of the *Retroviridae* family, the members of
which contain an RNA genome and reverse
transcriptase activity. During their growth
cycle, retroviruses copy their RNA into proviral
DNA. The proviral DNA is able to integrate into
25 the chromosomal DNA of the host cell where it uses
the transcriptional and translational machinery of
the host to express viral RNA and proteins.
Viruses are released from the cell by budding from
the cytoplasmic membrane. In the case of HIV-1
30 and HIV-2, viral replication results in the death
of helper T-cell host cells, which leads to a
state of severe immunodeficiency, to the
development of various malignancies and
opportunistic infections, and ultimately to the
35 death of the infected organism.

The incidence of AIDS has risen to epidemic proportions in many countries without the development of preventative treatments or therapies which are successful in the long term.

5 Those few therapeutic agents which have been prescribed, such as the nucleoside analogs 3'-azido-3'-deoxythymidine (AZT), dideoxyinosine (ddI), and dideoxycytosine (ddC), and various protease inhibitors have met with limited success.
10 This has been in part because of the cytotoxicity of these agents. In addition, some viruses escape due to mutations that render them insensitive to these agents and the difficulty of antiviral action due to the ability of the virus to
15 integrate into the host's genome. Thus, there is a long felt need for more effective therapeutic agents and preventative therapies for AIDS.

20 More recently new chemotherapeutic agents have been developed which are capable of modulating cellular and foreign gene expression. These agents, called antisense oligonucleotides, bind to a target single-stranded nucleic acid molecules according to the Watson-Crick or the
25 Hoogsteen rule of base pairing, and in doing so, disrupt the function of the target by one of several mechanisms: by preventing the binding of factors required for normal translation or transcription; in the case of an mRNA target, by
30 triggering the enzymatic destruction of the message by RNase H; or by destroying the target via reactive groups attached directly to the antisense oligonucleotide.

Antisense oligodeoxynucleotides have been designed to specifically inhibit the expression of HIV-1 and other viruses (see, e.g., Agrawal (1992) *Trends in Biotechnology* 10:152-158; Agrawal et al. in 5 *Gene Regulation: Biology of Antisense RNA and DNA* (Erickson and Izant, eds.) Raven Press Ltd., New York (1992) pp. 273-283); Matsukura et al. in *Prospects for Antisense Nucleic Acid Therapy of Cancer and AIDS*, Wiley-Liss, Inc. (1992) pp. 159-1798); and Agrawal (1991) in 10 *Prospects for Antisense Nucleic Acid Therapy for Cancer and AIDS*, (Wickstron, ed.) Liss, New York, pp. 145-148). For example, it has been shown that antisense oligonucleotides having phosphodiester internucleoside bonds and sequences complementary 15 to portions of genomic HIV-1 RNA inhibit viral replication in early infected cells (Zamecnik et al. (1986) *Proc. Acad. Sci. USA* 83:4143-4147; Goodchild et al. (1988) *Proc. Natl. Acad. Sci USA* 85:5507-5511).

20 However, these phosphodiester-linked molecules are less able to inhibit viral replication in chronically infected cells (Agrawal et al. (1989) *Proc. Natl. Acad. Sci USA* 86:7790-7794), mainly because of their nuclease susceptibility 25 (Wickstrom (1986) *J. Biochem. Biophys. Meth.* 13:97-102). Therefore, chemically modified, nuclease-resistant analogs have been developed which are effective in inhibiting HIV-1 replication in tissue cultures (see, Sarin et al. (1988) *Proc. Natl Acad. Sci. USA* 30 85:7448-7451; Agrawal et al. (1988) *Proc. Natl Acad. Sci USA* 85:7079-7083; Matsukura et al. (1988) *Gene* 72:343-347). These analogs include

oligonucleotides with nuclease-resistant
phosphorothioate internucleotide linkages shown to
inhibit HIV-1 replication in both acute infection
(U.S.S.N. 08/309,823; Agrawal et al. (1989) *Proc.*
5 *Natl. Acad. Sci USA* 86:7790-7794) and in chronically
infected cell lines (Agrawal et al. (1991) in *Gene*
Regulation: Biology of Antisense RNA, eds. Erickson et al.
(Raven Press, New York), pp. 273-284; Vickers et
al. (1991) *Nucleic Acids Res.* 19:3359-3368; Matsukura
10 et al. (1989) *Proc. Natl Acad. Sci.* 86:4244-4248;
Agrawal et al. (1988) *Proc. Natl Acad. Sci USA* 85:7079-
7083).

However, some phosphorothioate-linked
15 oligonucleotides that have "GC-rich" nucleotide
sequences have been found to evoke
immunostimulatory responses in the organisms to
whom they have been administered. For example,
Knierp et al. (*Nature* (1995) 374:546-549) discloses
20 that oligonucleotides containing the CG
dinucleotide flanked by certain other sequences
have a mitogenic and other side effects.

25 *SUB*
B1 Thus, there still remains a need for a more
effective anti-HIV oligonucleotide having
therapeutic effects that are accompanied fewer
side effects, e.g., little cellular toxicity and
reduced immunostimulatory response.

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SUMMARY OF THE INVENTION

5 It has been discovered that synthetic
oligonucleotides directed to a region of the HIV
gag inhibit HIV-1 and HIV-2 infection of mammalian
cells. These discoveries have been exploited to
develop the present invention, which in its
broadest aspect, provides synthetic
oligonucleotides having a nucleotide sequence
10 specifically complementary to nucleotides 325 to
346 of a conserved gag region of the HIV genome
set forth as SEQ ID NO:3. These oligonucleotides
have 21 nucleotides ("21mers") which are linked
via phosphorothioate internucleotide linkages.
15 Such phosphorothioate linkages contain a
substitution of sulfur for oxygen, thereby
rendering the oligonucleotide resistant to
nucleolytic degradation. The phosphorothioate
linkages may be mixed R_p and S_p enantiomers, or
20 they may be stereoregular or substantially
stereoregular in either R_p or S_p form (see Iyer et
al. (1995) *Tetrahedron Asymmetry* 6:1051-1054).

As used herein, the term "synthetic
25 oligonucleotide" includes chemically synthesized
polymers of 12 to 50, preferably from about 15 to
about 30, and most preferably, 21 ribonucleotide
and/or deoxyribonucleotide monomers connected
together or linked by at least one, and preferably
30 more than one, 5' to 3' internucleotide linkage.
The term "nucleotide sequence specifically
complementary to" nucleotides 324 to 345 of a
conserved gag region of the HIV genome is intended
to mean a sequence of nucleotides that binds to

the *gag* genomic RNA, proviral DNA, or mRNA
sequence under physiological conditions, e.g., by
Watson-Crick base pairing (interaction between
oligonucleotide and single-stranded nucleic acid)
5 or by Hoogsteen base pairing (interaction between
oligonucleotide and double-stranded nucleic acid)
or by any other means including in the case of a
oligonucleotide binding to RNA, causing pseudoknot
formation. Binding by Watson-Crick or Hoogsteen
10 base pairing under physiological conditions is
measured as a practical matter by observing
interference with the function of the nucleic acid
sequence. The term "a conserved *gag* region"
refers to a sequence of nucleotides within the
15 *gag* gene which is found in related HIV strains.

In one embodiment, the oligonucleotides of
the invention comprise at least two 3'-terminal
ribonucleotides, at least two 5'-terminal
20 ribonucleotides, or at least two 3'-terminal and
at least two 5' terminal ribonucleotides. In
preferred embodiments according to this aspect of
the invention, the oligonucleotide is a core
region hybrid oligonucleotide comprising a region
25 of at least two deoxyribonucleotides, flanked by
5' and 3' ribonucleotide regions, each having at
least two ribonucleotides. In one particular
embodiment, the oligonucleotides of the invention
have four contiguous 3'-terminal ribonucleotides
30 and four contiguous 3'-terminal ribonucleotides,
flanking 13 deoxynucleotides.

In preferred embodiments, the ribonucleotides
in the hybrid oligonucleotide are 2'-substituted

ribonucleotides. For purposes of the invention,
the term "2'-substituted" means substitution of
the 2' position of the pentose moiety with an -O-
lower alkyl group containing one to six saturated
5 or unsaturated carbon atoms, or with an -O-aryl or
allyl group having two to six carbon atoms,
wherein such alkyl, aryl or allyl group may be
unsubstituted or may be substituted, e.g., with
halo, hydroxy, trifluoromethyl, cyano, nitro,
10 acyl, acyloxy, alkoxy, carboxyl, carbalkoxyl, or
amino groups; or with a hydroxy, an amino or a
halo group, but not with a 2'-H group. In
specific embodiments, the ribonucleotides are 2'-
O-alkyl ribonucleotides such as 2'-O-methyl
15 ribonucleotides.

In particular embodiments, the
oligonucleotides of the invention have SEQ ID
NO:1, NO:2, NO:3, or NO:4. In some embodiments,
20 these oligonucleotides inhibit HIV-1 or HIV-2
infection in a cell and/or exhibit antiviral
activity against HIV-1 and HIV-2.

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25 In yet another aspect, the invention provides
pharmaceutical formulations suitable for
inhibiting and treating HIV-1 or HIV-2 infection
and having reduced side effects such as
immunogenicity. These formulations and for
inhibiting comprising at least one oligonucleotide
30 in accordance with the invention in a
pharmaceutically acceptable carrier.

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As used herein, a "pharmaceutically or
physiologically acceptable carrier" includes any

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and all solvents (including but limited to lactose), dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions of the invention is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

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In another aspect, the invention provides a method of treating HIV-1 or HIV-2 infection in a mammal. In this method an oligonucleotide according to the invention is administered to the mammal in an amount effective to inhibit the proliferation of the virus. For purposes of the invention, the term "mammal" is meant to encompass primates and humans. In some embodiments, the oligonucleotide is orally administered to the mammal. The term "orally administered" refers to the provision of the formulation via the mouth through ingestion, or via some other part of the gastrointestinal system including the esophagus. In other embodiments, the oligonucleotide is administered via intravenous injection. In yet other embodiments, the oligonucleotide is administered colorectally. The term "colorectal administration" or "rectal administration" or "colorectally administered" refers to the provision of the pharmaceutical formulation of the invention to any part of the large intestine via

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surgical implantation, anal administration, or any other mode of placement therein.

5 The invention also provides in another aspect a method of inhibiting HIV-1 or HIV-2 infection in a cell. In this method the cell is contacted with a synthetic oligonucleotide according to the invention.

10 In yet another aspect, the invention provides a method for introducing an intact oligonucleotide into a mammal. This method comprises administering to the mammal an oligonucleotide according to the invention which is present in
15 intact form in the systemic plasma of the mammal following oral administration. In one embodiment, the oligonucleotide is orally or enterally administered. In another embodiment, the
20 oligonucleotide is intravenously administered.

BRIEF DESCRIPTION OF THE DRAWINGS

The foregoing and other objects of the present invention, the various features thereof,
5 as well as the invention itself may be more fully understood from the following description, when read together with the accompanying drawings in which:

10 FIG. 1 is a graphic representation of the inhibition of HIV-1 infection in cells treated during initial infection with a 4X4 hybrid oligonucleotide of the invention having SEQ ID
15 NO:1;

FIG. 2 is a graphic representation of the inhibition of HIV-1 infection in cells treated following initial infection with a 4X4 hybrid oligonucleotide of the invention having SEQ ID
20 NO:1; and

FIG. 3 is a graphic representation of the results of an XTT assay demonstrating the ability
25 of a 4x4 oligonucleotide of the invention having SEQ ID NO:1 to inhibit HIV-2-induced cell killing.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

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5 The patent and scientific literature referred to herein establishes the knowledge that is available to those with skill in the art. The issued U.S. patent, allowed patent applications, and articles cited herein are hereby incorporated by reference.

10 It is known that antisense oligonucleotides, can bind to a target single-stranded nucleic acid molecule according to the Watson-Crick or the Hoogsteen rule of base pairing, and in doing so, disrupt the function of the target by one of
15 several mechanisms: by preventing the binding of factors required for normal transcription, splicing, or translation; by triggering the enzymatic destruction of mRNA by RNase H if a contiguous region of deoxyribonucleotides exists
20 in the oligonucleotide, and/or by destroying the target via reactive groups attached directly to the antisense oligonucleotide.

25 Novel antisense oligonucleotides have been designed which inhibit HIV-1 and HIV-2 replication. These oligonucleotides are synthetic oligonucleotides having phosphorothioate internucleotide linkages and a nucleotide sequence that is complementary to a portion of the *gag*
30 region of the genome of HIV-1 and HIV-2. Sequences situated in this region have been demonstrated to be essential for viral packaging. These sequences form a stable secondary structure (Harrison et al. (1991) in *RNA Tumor Viruses* (Coffin

et al., eds.) Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, pp. 235). The oligonucleotides of the invention have been designed to bind to this region of RNA and DNA, thereby disrupting its natural stability and resulting ultimately in the inhibition of viral packaging and translation of *gag* mRNA. The specific sequence to which the oligonucleotides of the invention are complementary is nucleotides 324-345 of the *gag* region of HIV-1. This sequence is very conserved among strains of HIV-1, as shown below in TABLE 1.

TABLE 1

15	Sequence of:		
	324-345→	TCTTCCTCTCTTACCCACGCT	
	CONSENSUS	CGGAGGCTAGAAGGAGAGAGATGGGTGCGAGAGCGTCAGTA	
	Strains	.	.
20	<u>of HIV-1</u>	.	.
	HTLV/LLAV	G	A
	HIVLAI	G	A
	HIVNL43	G	G
	HIVMN	G	G
25	HIVJH3	G	A
	HIVYOI	G	A
	HIVCDC4	G	A
	HIVRF	G	A
	HIVMAL	G	A
30			(African)
	HIVU455	A	A CCTCAG
			(Ugandan)
35	HIVSF2	(GA) 4G	G
	HIVNDK	G	A

Targeting an antisense oligonucleotide to such a conserved region including an active gene allows for efficient inhibition of HIV proliferation without the generation of "escape mutants." Escape mutants arise when a mutation occurs in a region of the genome targeted by the antisense oligonucleotide. They occur at a higher frequency in non-coding regions (like the SA region of HIV-1) than in regions encoding a protein.

Oligonucleotides of the invention are more specific, less toxic, and have greater nuclease resistance than many other chemotherapeutic agents designed to inhibit HIV replication. In particular, these oligonucleotides are less immunostimulatory than other oligonucleotides directed to the HIV-1 *gag* sequence because their nucleotide sequences are not GC-rich. Furthermore, these hybrid oligonucleotides having phosphorothioate linkages are more resistant to nucleolytic degradation than are DNA compounds having solely phosphodiester linkages.

The oligonucleotides useful in the method of the invention are at least 12 nucleotides in length, but are preferably 15 to 21 nucleotides long, with 21mers being most common. They are composed of deoxyribonucleotides, ribonucleotides, or a combination of both (i.e., are "hybrids"), with the 5' end of one nucleotide and the 3' end of another nucleotide being covalently linked by phosphorodithioates or phosphorothioates, non-

phosphodiester internucleotide linkages.
Oligonucleotides with these linkages can be
prepared according to known methods such as
phosphoramidate or H-phosphonate chemistry which
5 can be carried out manually or by an automated
synthesizer as described by Brown (*A Brief History of
Oligonucleotide Synthesis. Protocols for Oligonucleotides and Analogs,
Methods in Molecular Biology* (1994) 20:1-8). (See also,
e.g., Sonveaux "Protecting Groups in
10 Oligonucleotides Synthesis" in Agrawal (1994)
Methods in Molecular Biology 26:1-72; Uhlmann et al.
(1990) *Chem. Rev.* 90:543-583).

The oligonucleotides of the composition may
15 also be additionally modified in a number of ways
without compromising their ability to hybridize to
the target nucleic acid. Such modifications
include, for example, those which are internal or
at the end(s) of the oligonucleotide molecule and
20 include additions to the molecule of the
internucleoside phosphate linkages, such as
cholesteryl or diamine compounds with varying
numbers of carbon residues between the amino
groups and terminal ribose, deoxyribose and
25 phosphate modifications which cleave, or crosslink
to the opposite chains or to associated enzymes or
other proteins which bind to the viral genome.
Examples of such modified oligonucleotides include
oligonucleotides with a modified base and/or sugar
30 such as arabinose instead of ribose, or a 3', 5'-
substituted oligonucleotide having a sugar which,
at both its 3' and 5' positions is attached to a
chemical group other than a hydroxyl group (at its

3' position) and other than a phosphate group (at its 5' position). Other modified oligonucleotides are capped with a nuclease resistance-conferring bulky substituent at their 3' and/or 5' end(s) ,
5 or have a substitution in one nonbridging oxygen per nucleotide. Such modifications can be at some or all of the internucleoside linkages, as well as at either or both ends of the oligonucleotide and/or in the interior of the molecule. For the
10 preparation of such modified oligonucleotides, see, e.g., Agrawal (1994) *Methods in Molecular Biology* 26; Uhlmann et al. (1990) *Chem. Rev.* 90:543-583). Oligonucleotides which are self-stabilized are also considered to be modified oligonucleotides
15 useful in the methods of the invention (Tang et al. (1993) *Nucleic Acids Res.* 20:2729-2735). These oligonucleotides comprise two regions: a target hybridizing region; and a self-complementary region having an oligonucleotide sequence
20 complementary to a nucleic acid sequence that is within the self-stabilized oligonucleotide.

The preparation of these unmodified and modified oligonucleotides is well known in the art
25 (reviewed in Agrawal et al. (1992) *Trends Biotechnol.* 10:152-158; see, e.g., Uhlmann et al. (1990) *Chem. Rev.* 90:543-584; and (1987) *Tetrahedron. Lett.* 28:(31):3539-3542); Agrawal (1994) *Methods in Molecular Biology* 20:63-80); and Zhang et al. (1996) *J. Pharmacol. Expt. Thera.*
30 278:1-5).

Preferred oligonucleotides according to the invention are hybrid oligonucleotides in that they

contain both deoxyribonucleotides and at least two
2' substituted ribonucleotides at their
termin(i/us). For purposes of the invention, the
term "2'-substituted" means substitution at the 2'
5 position of the ribose with, e.g., a -O-lower
alkyl containing 1-6 carbon atoms, aryl or
substituted aryl or allyl having 2-6 carbon atoms
e.g., 2'-O-allyl, 2'-O-aryl, 2'-O-alkyl, 2'-halo,
or 2'-amino, but not with 2'-H, wherein allyl,
10 aryl, or alkyl groups may be unsubstituted or
substituted, e.g., with halo, hydroxy,
trifluoromethyl, cyano, nitro, acyl, acyloxy,
alkoxy, carboxyl, carbalkoxyl or amino groups.
Useful substituted ribonucleotides are 2'-O-alkyls
15 such as 2'-O-methyl, 2'-O-ethyl, and 2'-O-propyl,
with 2'-O-methyl being the most preferred.

The hybrid oligonucleotides useful in the
method of the invention resist nucleolytic
20 degradation, form stable duplexes with RNA or DNA,
and preferably activate RNase H when hybridized
with RNA. They may additionally include at least
one unsubstituted ribonucleotide. For example, an
oligonucleotide useful in the method of the
25 invention may contain all deoxyribonucleotides
with the exception of two 2' substituted
ribonucleotides at the 3' terminus of the
oligonucleotide, or the 5' terminus of the
oligonucleotide. Alternatively, the
30 oligonucleotide may have at least two, and
preferably 4, substituted ribonucleotides at both
its 3' and 5' termini.

Preferred oligonucleotides have at least two and preferably four 2'-0-methyl ribonucleotides at both the 3' and 5' termini, with the remaining nucleotides being deoxyribonucleotides. One preferred oligonucleotide is a 21mer phosphorothioate linked oligonucleotide containing therein deoxyribonucleotides flanked on each side by four 2'-0-methyl ribonucleotides. This preferred oligonucleotide is referred to as a "4x4". One preferred class of oligonucleotides useful in the method of the invention contains four or more deoxyribonucleotides in a contiguous block, so as to provide an activating segment for RNase H. In certain cases, more than one such activating segment will be present at any location within the interior of the oligonucleotide. There may be a majority of deoxyribonucleotides in oligonucleotides according to the invention. In fact, such oligonucleotides may have as many as all but two nucleotide being deoxyribonucleotides.

TABLE 2 lists some representative species of oligonucleotides which are useful in the method of the invention. 2'-substituted nucleotides are underscored.

TABLE 2

30	OLIGO NO.	OLIGONUCLEOTIDE (5'→3')	SEQ ID NO:
	1	UCGCACCCATCTCTCTCCUUC	1
	2	UCGCACCCATCTCTCTCCU <u>UC</u>	1
35	3	UCGCACCCATCTCTCTCC <u>UUC</u>	1
	4	UCGCACCCATCTCTCTCC <u>UUC</u>	1

TABLE 2 (cont.)

5	5	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	6	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	7	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	8	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	9	<u>UCGCACCCATCTCTCTCCUUC</u>	1
10	10	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	11	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	12	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	13	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	14	<u>UCGCACCCATCTCTCTCCUUC</u>	1
15	15	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	16	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	17	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	18	<u>UCGCACCCATCTCTCTCCUUC</u>	1
	19	<u>UCGCACCCATCTCTCTCCUUC</u>	1
20	20	TCGCACCCATCTCTCTCCTTC	2
	21	CGCACCCATCTCTCTCCUUCU	3
	22	CGCACCCATCTCTCTCCUUCU	3
	23	CGCACCCATCTCTCTCCUUCU	3
	24	CGCACCCATCTCTCTCCUUCU	3
25	25	CGCACCCATCTCTCTCCUUCU	3
	26	CGCACCCATCTCTCTCCUUCU	3
	27	CGCACCCATCTCTCTCCUUCU	3
	28	CGCACCCATCTCTCTCCUUCU	3
	29	CGCACCCATCTCTCTCCUUCU	3
30	30	CGCACCCATCTCTCTCCUUCU	3
	31	CGCACCCATCTCTCTCCUUCU	3
	32	CGCACCCATCTCTCTCCUUCU	3
	33	CGCACCCATCTCTCTCCUUCU	3
	34	CGCACCCATCTCTCTCCUUCU	3
	35	CGCACCCATCTCTCTCCUUCU	3
	36	CGCACCCATCTCTCTCCUUCU	3

TABLE 2 (cont.)

5	37	<u>CGCACCCATCTCTCTCCUUCU</u>	3
	38	<u>CGCACCCATCTCTCTCCUUCU</u>	3
	39	<u>CGCACCCATCTCTCTCCUUCU</u>	3
	40	CGCACCCATCTCTCTCCTTCT	4

10 Oligonucleotides as described above are
useful in a method of inhibiting HIV-1 or HIV-2
infection in a cell. In this method a cell is
contacted with an oligonucleotide of the invention
such that virus present in the cell at the time of
15 contact, or after such contact is unable to
replicate.

 To determine whether oligonucleotides of the
invention could inhibit or prevent HIV infection,
20 cytopathic effect- (CPE-)based infection
experiments were performed in MT-4 cells. The
results of these studies indicate that
oligonucleotides of the invention can both inhibit
an existing infection (FIG. 1) and protect against
25 such infection (FIG. 2).

 In addition, it was determined that synthetic
oligonucleotides systemically administered to
pregnant murine females crossed the placenta and
30 became available in the blood of embryos in utero.
Thus, it is contemplated that oligonucleotides of
the invention will be used in a method of treating
the fetuses and human mothers harboring HIV.

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The oligonucleotides described herein are administered to the mammal in the form of therapeutic pharmaceutical formulations that are effective for treating virus infection. These pharmaceutical formulation may be administered in conjunction with other therapeutic agents, e.g., AZT and/or various protease inhibitors, to treat AIDS.

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The therapeutic pharmaceutical formulation containing at least one oligonucleotide according to the invention includes a physiologically acceptable carrier which is congruent with the mode of administration. Examples include an inert diluent or an assimilable edible carrier.

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Suitable formulations that include pharmaceutically acceptable excipients for introducing compounds to the bloodstream by intravenous injection and other than injection routes can be found in *Remington's Pharmaceutical Sciences* (18th ed.) (Genarro, ed. (1990) Mack Publishing Co., Easton, PA).

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The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases the form must be sterile. It must be stable under the

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conditions of manufacture and storage and may be preserved against the contaminating action of microorganisms, such as bacterial and fungi. The carrier can be a solvent or dispersion medium. The prevention of the action of microorganisms can

be brought about by various antibacterial and antifungal agents. Prolonged absorption of the injectable therapeutic agents can be brought about by the use of the compositions of agents delaying absorption. Sterile injectable solutions are prepared by incorporating the oligonucleotide in the required amount in the appropriate solvent, followed by filtered sterilization.

Alternatively, the oligonucleotide of the invention and other ingredients may be enclosed in a hard or soft shell gelatin capsule, compressed into tablets, or incorporated directly into the individual's diet. The oligonucleotide may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. When the oligonucleotide is administered orally, it may be mixed with other food forms and pharmaceutically acceptable flavor enhancers. When the oligonucleotide is administered enterally, they may be introduced in a solid, semi-solid, suspension, or emulsion form and may be compounded with any number of well-known, pharmaceutically acceptable additives. Sustained release oral delivery systems and/or enteric coatings for orally administered dosage forms are also contemplated such as those described in U.S. Patent Nos. 4,704,295, 4,556,552, 4,309,404, and 4,309,406.

As used herein, the term "therapeutically effective amount" means the total amount of each active component of the pharmaceutical formulation

or method that is sufficient to show a meaningful subject or patient benefit, i.e., a reduction in tumor growth or in the expression of proteins which cause or characterize the cancer. When applied to an individual active ingredient, administered alone, the term refers to that ingredient alone. When applied to a combination, the term refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously.

A "therapeutically effective manner" refers to a route, duration, and frequency of administration of the pharmaceutical formulation which ultimately results in meaningful patient benefit, as described above. In some embodiments of the invention, the pharmaceutical formulation is administered via injection, sublingually, colorectally, intradermally, orally, enterally or in bolus, continuous, intermittent, or continuous, followed by intermittent regimens.

The therapeutically effective amount of synthetic oligonucleotide administered in the method of the invention will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of synthetic oligonucleotide with which to treat each individual patient. Initially, the attending physician may administer low doses of the synthetic oligonucleotide and observe the

patient's response. Larger doses of synthetic oligonucleotide may be administered until the optimal therapeutic effect is obtained for the patient, and at that point the dosage is not increased further. It is contemplated that the dosages of the pharmaceutical compositions administered in the method of the present invention should contain about 0.1 to 100.0 mg/kg body weight per day, preferably 0.1 to 75.0 mg/kg body weight per day, more preferably, 1.0 to 50.0 mg/kg body weight per day, even more preferably, 1 to 25 mg/kg body weight per day, and even more preferably, 1 to 10 or 1 to 5.0 mg/kg body weight per day. The oligonucleotide is preferably administered at a sufficient dosage to attain a blood level of oligonucleotide from about 0.01 μ M to about 100 μ M. Preferably, the concentration of oligonucleotide at the site of aberrant gene expression should be from about 0.01 μ M to about 50 μ M, more preferably, from about 0.01 μ M to about 10 μ M, and most preferably from about 0.05 μ M to about 5 μ M. However, for localized administration, much lower concentrations than this may be effective, and much higher concentrations may be tolerated. It may be desirable to administer simultaneously or sequentially a therapeutically effective amount of one or more of the therapeutic compositions of the invention when individual as a single treatment episode.

It will be appreciated that the unit content of active ingredient or ingredients contained in an individual dose of each dosage form need not in

itself constitute an effective amount since the necessary effective amount can be reached by administration of a plurality of dosage units (such as suppositories, gels, or creams, or combinations thereof). In fact, multi-dosing (once a day) has been shown to significantly increase the plasma and tissue concentrations of MBO's (data not shown).

10 ~~Administration of pharmaceutical compositions~~
in accordance with invention or to practice the
method of the present invention can be carried out
in a variety of conventional ways, such as by oral
ingestion, enteral, colorectal, or transdermal
15 administration, inhalation, sublingual
administration, or cutaneous, subcutaneous,
intramuscular, intraocular, intraperitoneal, or
intravenous injection, or any other route of
administration known in the art for administering
20 therapeutic agents.

When the composition is to be administered orally, sublingually, or by any non-injectable route, the therapeutic formulation will preferably include a physiologically acceptable carrier, such as an inert diluent or an assimilable edible carrier with which the composition is administered. Suitable formulations that include pharmaceutically acceptable excipients for introducing compounds to the bloodstream by other than injection routes can be found in *Remington's Pharmaceutical Sciences* (18th ed.) (Genarro, ed. (1990) Mack Publishing Co., Easton, PA). The oligonucleotide and other ingredients may be

enclosed in a hard or soft shell gelatin capsule,
compressed into tablets, or incorporated directly
into the individual's diet. The therapeutic
compositions may be incorporated with excipients
5 and used in the form of ingestible tablets, buccal
tablets, troches, capsules, elixirs, suspensions,
syrups, wafers, and the like. When the
therapeutic composition is administered orally, it
may be mixed with other food forms and
10 pharmaceutically acceptable flavor enhancers.
When the therapeutic composition is administered
enterally, they may be introduced in a solid,
semi-solid, suspension, or emulsion form and may
be compounded with any number of well-known,
15 pharmaceutically acceptable additives. Sustained
release oral delivery systems and/or enteric
coatings for orally administered dosage forms are
also contemplated such as those described in U.S.
Patent Nos. 4,704,295, 4,556,552, 4,309,404, and
20 4,309,406.

SUB
25 20
When a therapeutically effective amount of
composition of the invention is administered by
injection, the synthetic oligonucleotide will
preferably be in the form of a pyrogen-free,
parenterally-acceptable, aqueous solution. The
preparation of such parenterally-acceptable
solutions, having due regard to ph, isotonicity,
stability, and the like, is within the skill in
30 the art. A preferred pharmaceutical composition
for injection should contain, in addition to the
synthetic oligonucleotide, an isotonic vehicle
such as Sodium Chloride Injection, Ringer's
Injection, Dextrose Injection, Dextrose and Sodium

Chloride Injection, Lactated Ringer's Injection,
or other vehicle as known in the art. The
pharmaceutical composition of the present
invention may also contain stabilizers,
5 preservatives, buffers, antioxidants, or other
additives known to those of skill in the art. The
pharmaceutical formulation can be administered in
bolus, continuous, or intermittent dosages, or in
a combination of continuous and intermittent
10 dosages, as determined by the physician and the
degree and/or stage of illness of the patient.
The duration of therapy using the pharmaceutical
composition of the present invention will vary,
depending on the unique characteristics of the
15 oligonucleotide and the particular therapeutic
effect to be achieved, the limitations inherent in
the art of preparing such a therapeutic
formulation for the treatment of humans, the
severity of the disease being treated and the
20 condition and potential idiosyncratic response of
each individual patient. Ultimately the attending
physician will decide on the appropriate duration
of intravenous therapy using the pharmaceutical
composition of the present invention.

25

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30

To determine the preclinical range of anti-
HIV activity of various oligonucleotides of the
invention (see TABLE 2), evaluations were
performed with Oligo 12 (having SEQ ID NO:1),
Oligo 32 (SEQ ID NO:3) and Oligo 41 (SEQ ID NO:6).
These evaluations were performed to determine the
activity of these compounds against a variety of
wild type and drug-resistant strains of HIV-1,
including both laboratory derived and low passage,

5
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 B₉

clinical strains of virus and T-lymphocyte-tropic and monocyte-macrophage-tropic viruses same as these are listed below in TABLE 3.

TABLE 3

BIOLOGICAL PROPERTIES OF CLINICAL STRAINS OF HIV-1

	ISOLATE	TROPISM	AZT IC ₅₀ (μM)	ddi IC ₅₀ (μM)	SYNCYTIA	GROWTH
	BAKI	L	0.049	2.61	SI	R/H
10	SLKA	M	0.025	0.32	NSI	S/L
	WEJO	L	0.056	2.18	SI	R/H
	ROJO	L	0.016	0.87	SI	R/H
	ROMA	M	0.016	0.16	SI	R/H
	STDA	L	0.017	0.23	SI	R/H
15	WOME	L	0.016	0.41	SI	R/H
	VIHU	L	0.016	1.21	NSI	S/L
	TEKI	L	0.029	0.37	NSI	S/L
	TEKI	M	0.016	1.70	NSI	S/L
	DEJO	L	0.015	ND	NSI	S/L
20	BLCH	L	0.010	ND	NSI	S/L
	RIARL	L	0.010	ND	NSI	S/L

25 L - lymphocyte
 M - macrophage
~~SI - syncytium inducing~~
 NSI - non-syncytium inducing
 R/H - rapid/high
 S/L - slow/low

30

35 In addition, the activity of the compounds was evaluated against HIV-2, and the toxicity of Oligo 41 was evaluated by a variety of methods in infected and uninfected, established and fresh human cells.

5 The initial experiment performed involved
evaluation of Oligos 12, 32, and 41 against three
laboratory strains of HIV-1 (IIIIB, RF and SK1) and
one strain of HIV-2 (ROD) in parallel with the
positive control compound ddC in the XTT-based
anti-HIV assay. All these oligonucleotides are
active against both HIV-1 and HIV-2. An enhanced
level of activity was detected with these
compounds when evaluated against the HIV-2 strain
10 ROD. Representative results are shown in FIG. 3.

15 In another experiment, the anti-HIV activity
of Oligos 12, 32, and 41 was evaluated against a
variety of low passage, clinical strains of HIV-1
in fresh human peripheral blood mononuclear cells.
These strains include viruses obtained from
pediatric patients attending the Children's
Hospital University of Alabama at Birmingham as
well as viruses representative of the various HIV-
20 1 clades found throughout the world, shown below
in TABLE 4.

TABLE 4
BIOLOGICAL PROPERTIES OF CLADE VIRUS ISOLATES

	<u>VIRUS</u>	<u>CLADE</u>	<u>PHENOTYPE</u>	<u>COUNTRY</u>
	92RW009A	A	NSI	Rwanda
5	92UG029A	A	SI	Uganda
	92BR021B	B	SI	Brazil
	92TH026B	B	NSI	Thailand
	92BR025C	C	NSI	Brazil
	92UG021D	D	SI	Uganda
10	92UG035D	D	NSI	Uganda
	92TH022E	E	NSI	Thailand
	93BR029F	F	NSI	Brazil
	93BR020F	F	SI	Brazil

15

In addition to these T-tropic strains, the activity of the compounds was also evaluated against the monocyte-macrophage strains BaL and ADA. Oligos 12 and 32 according to the invention, as well as Oligo 41 are active against low passage clinical T-tropic strains of HIV-1. The activity of the compounds varies from strain to strain. The compounds were not active against the monocyte-macrophage-tropic strains BaL and ADA.

25

In other studies, the anti-HIV activity of Oligos 12, 32, and 41 was evaluated against a variety of drug-resistant virus strains, including viruses resistant to nevirapine (N119), 3TC (M198I), protease inhibitors (JE105/R and KN1272/R) and AZT (4xAZT-R).

30

The results of these evaluations indicate that these oligonucleotides remained active against viruses resistant to nevirapine, 3TC and the protease inhibitors, but were less active against viruses with mutations conferring resistance to AZT. An enhanced level of activity was detected against the nevirapine-resistant strain N119.

In yet another experiment, the toxicity of Oligo 41 was evaluated in uninfected and HIV-1 infected fresh human peripheral blood mononuclear cells, using a variety of quantitative endpoints. Toxicity was evaluated using the tetrazolium dyes XTT or MTT, trypan blue cell and cell viability counting and the incorporation of tritiated thymidine. Two replicate assays were performed. In the first assay, Oligo 41 was used at a high test concentration of 50 µg/ml and toxicity was evaluated on day 7. No toxicity was detected by any of the quantitative endpoints employed. A second assay was performed to further evaluate toxicity at higher compound concentration and with longer exposure to the compound. In this assay, employing a high test concentration of 150 µg/ml and extending the time of drug exposure from 7 days until 14 days, once again no toxicity was detected.

30 In another set of experiments, the bioavailability of Oligo 12 was examined *in vivo* was found to be intravenously and orally bioavailable to rats and monkeys after a single dose.

In addition, synthetic oligonucleotides systemically administered to pregnant murine females were found to cross the placenta and be available in the blood of embryos *in utero*. Thus, it is contemplated that oligonucleotides of the invention be used in methods of treating human fetuses and mothers harboring HIV.

In order to determine if the oligonucleotide of the invention administered according to the method of the invention is absorbed into body tissues, and if so, in which tissues absorption occurs, the following study was performed. Samples of various body tissues from treated monkeys and rats were analyzed for radioactivity at increasing hours after intravenous or oral administration of a radioactively labelled oligonucleotide specific for HIV. This oligonucleotide was found to be absorbed through the gastrointestinal tract and accumulated in various organs and tissues.

To evaluate the chemical form of radioactivity in plasma HPLC is used to demonstrate the presence of both intact oligonucleotide as well as metabolites various hours after oral administration. Intact oligonucleotide may also be detected in liver various hours after administration. Further evidence to support the absorption of the oligonucleotide may come from urine sample analysis after radioactively labelled gag-specific oligonucleotide was orally administered. That the oligonucleotide continues to be excreted in the

urine over time following the administration of radiolabelled oligonucleotide implies that other tissues had absorbed it, and that the body was capable of absorption for an extended period of time.

The following examples illustrate the preferred modes of making and practicing the present invention, but are not meant to limit the scope of the invention since alternative methods may be utilized to obtain similar results.

EXAMPLE 1

Synthesis and Purification of Oligonucleotides

Oligonucleotide phosphorothioates were synthesized using an automated DNA synthesizer (Model 8700, Biosearch, Bedford, MA) using a beta-cyanoethyl phosphoramidate approach on a 10 micromole scale. To generate the phosphorothioate linkages, the intermediate phosphite linkage obtained after each coupling was oxidized using 3H, 1,2-benzodithiole-3H-one-1,1-dioxide (see Beaucage, in *Protocols for Oligonucleotides and Analogs: Synthesis and Properties*, Agrawal (ed.), (1993) Humana Press, Totowa, NJ, pp. 33-62).

Hybrid oligonucleotides were synthesized similarly, except that segments containing 2'-O-methylribonucleotides were assembled using 2'-O-methylribonucleoside phosphoramidite, followed by oxidation to a phosphorothioate or phosphodiester linkage as described above. Deprotection and purification of oligonucleotides was carried out

according to standard procedures, (see Padmapriya et al. (1994) *Antisense Res. & Dev.* 4:185-199).

5

EXAMPLE 2

Propagation and Quantitation of Cell Lines and Virus Stocks

A. Cells

10

The CEM-SS cell line (Southern Research Institute-Frederick Research Center, Frederick, MD) is highly susceptible to infection with HIV, rapidly form multinucleated syncytia, and are eventually killed by HIV. The cells were maintained ($2-7 \times 10^5$ cells per ml) in RPMI 1640 tissue culture medium supplemented with 10% fetal bovine serum, glutamine, and antibiotics, and were passed twice weekly at 1:20 dilution. Passage number was logged each week. Cells were discarded after twenty weeks of passage and fresh CEM-SS cells thawed and utilized in the assay. Stocks of CEM-SS cells were frozen in liquid nitrogen in 1 ml NUNC vials in 90% fetal calf serum and 10% dimethyl sulfoxide (DMSO). Following thawing, CEM-SS cells were routinely ready to be utilized in the primary screen assay after two weeks in culture. Prior to replacing a late passage cell line, the new CEM-SS cells were tested in the screening assay protocol utilizing the current stock of infectious virus and AZT. If the infectivity of the virus was significantly different on the new cells or if AZT appeared less active than expected the new cells were not

25

30

entered into the screening program. Mycoplasma testing was routinely performed on all cell lines.

5 Other viral isolates tested included the following drug resistant strains.

10 The N119 isolate was derived *in vitro* by culture of the clinical strain A018 in the presence of the nonnucleoside reverse transcriptase inhibitor nevirapine. This isolate was obtained from the NIAID AIDS research and Reference Reagent Program (catalog #1392). The isolate possesses one mutation in the reverse transcriptase (Y181C) and we have found the
15 isolate to be extremely cytopathic to T cells such as CEM-SS and MT2 (Richman et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:11241). The 3TC/M1841 isolate was selected *in vitro* using the wild type IIIB strain of virus and sequential passage of the virus in
20 the presence of increasing drug concentration in CEM-SS cells (Buckheit Jr. et al. (1996) *Antimicrob. Chem. Chemother.* 7:243-252). The JE 105/R isolate was derived from sequential passage of IIIB in the presence of a protease inhibitor. This isolate
25 possesses the I84V and S37N amino acid changes in the protease. The KNI272/R isolate was derived from sequential passage of IIIB in the presence of the protease inhibitor KNI 272. The isolate possesses three amino acid changes in the
30 protease, F53L, A71V and T80I. The 4xAZT-Ri isolate was obtained by site-directed mutagenesis through introduction of four amino acid changes in the reverse transcriptase of the NL4-3 wild type

virus. The four amino acid changes are D67N, K70R, T215Y, and K219Q.

B. Virus

5

Virus pools (Southern Research Institute-Frederick Research Center, Frederick M) were prepared and titrated in CEM-SS cells, placed in 5 ml aliquots, and frozen at -135°C. After thawing, unused virus is discarded to avoid changes in infectious titer. Virus pools were prepared by the acute infection of 5×10^5 CEM-SS cells with HIV in a volume of 200 μ l at a multiplicity of infection determined to give complete cell killing at day 7 post-infection (approximately 0.05 for the III_B isolate of HIV-1 and 0.01 for the RF isolate of HIV-1).

10

15

C. Assay

20

Infection was allowed to proceed for one hour at 37°C, after which the cells were transferred to a T25 flask and the volume increased to 2 ml. On day 1 post-infection the volume was brought to 5 ml and on day 2 the volume was increased to 10 ml. Beginning on day 4, the cells were pelleted, the supernatant saved, and the cells resuspended in a fresh 10 ml aliquot of tissue culture medium. Complete medium changes on a daily basis, rather than allowing growth of the cells in the medium for longer periods of time, allowed the virus inoculum utilized in the primary screen to remain relatively undepleted of nutrients when it is used to infect cells. The staining reaction utilized

25

30

(XTT, see method below) required that the glucose concentration remain high (161). Wells depleted of glucose by cell growth will not permit metabolic conversion of the tetrazolium dye to the formazan product.

Cell-free supernatants from the acutely infected cells were saved on day 4, day 5, day 6, and day 7. An aliquot of supernatant was saved separately on each day for use in titer determination. Titer determinations included reverse transcriptase activity assay (see below), endpoint titration or plaque assay (CEM-SS) quantification of infectious particles (see below), and quantification of cell killing kinetics.

It has been determined that peak levels of infectious virus are produced in the acutely infected cultures as the viability of the cells falls through the 50% level. Since the primary screening assay quantifies the protective effects of a compound by its ability to inhibit HIV-induced cytopathic effects, the quantity of virus required to kill CEM-SS cells in 6 days was routinely utilized to determine the amount of virus required per well in the primary screening assay. Each of the daily pools was titrated in the primary screening tetrazolium dye XTT assay protocol (see below) by performing two-fold dilutions of the virus beginning at a high test concentration of 50 μ l of virus per well. The XTT staining method was utilized to determine the exact amount of virus required to kill all of the

CEM-SS cells in each well and this minimum amount of virus was utilized for performance of all primary testing. Identical methods were utilized to prepare all virus isolates utilized, including laboratory-derived strains of HIV-1, HIV-2 and SIV. Clinical isolates utilized were passaged in fresh human cells. The methods for the growth of these cells and the production of virus pools is described below.

EXAMPLE 3

Microtiter Antiviral XTT Assay

A. Cell Preparation:

CEM-SS cells (AIDS Research and Reference Reagent Program, NIH) or other established human T cell lines used in these experiments were passaged in T-150 flasks for use in the assay. On the day preceding the assay, the cells were split 1:2 to assure they would be in an exponential growth phase at time of infection. On the day of assay the cells were washed twice with tissue culture medium and resuspended in fresh tissue culture medium. Total cell and viability counting was performed using a hemacytometer and trypan blue dye exclusion. Cell viability was greater than 95% for the cells to be utilized in the assay. The cells were pelleted and resuspended at 2.5×10^4 cells per ml in tissue culture medium. Cells were added to the drug-containing plates in a volume of 50 μ l.

B. Virus Preparation

5 A pretitered aliquot of virus was removed from the freezer -80°C) and allowed to thaw slowly to room temperature in a biological safety cabinet. The virus was resuspended and diluted into tissue culture medium such that the amount of virus added to each well in a volume of $50\ \mu\text{l}$ will be the amount determined to give complete cell killing at 6 days post-infection. In general the virus pools produced with the IIIB isolate of HIV required the addition of $5\ \mu\text{l}$ of virus per well. Pools of RF virus were five to ten-fold more potent, requiring 0.5 to $1\ \mu\text{l}$ per well. TCID_{50} calculation by endpoint titration in CEM-SS cells indicated that the multiplicity of infection of these assays ranged from 0.005 to 2.5 .

C. Plate Format

20 Each plate contained cell control wells (cells only), virus control wells (cells plus virus), drug toxicity control wells (cells plus drug only), drug colorimetric control wells (drug only) as well as experimental wells (drug plus cells plus virus).

D. XTT Staining of Screening Plates

30 After 6 days of incubation at 37°C in a 5% CO_2 incubator, the test plates were analyzed by staining with the tetrazolium dye XTT. XTT-tetrazolium is metabolized by the mitochondrial enzymes of metabolically active cells to a soluble

formazan product, allowing the rapid quantitative analysis of the inhibition of HIV-induced cell killing by anti-HIV test substances. On day 6 post-infection plates were removed from the incubator and observed. The use of round bottom microtiter plates allows rapid macroscopic analysis of the activity of a given test compound by the evaluation of pellet size. The results of the macroscopic observations were confirmed and enhanced by further microscopic analysis.

XTT solution was prepared daily as a stock of 1 mg/ml in PBS. Phenazine methosulfate (PMS) solution was prepared at 15 mg/ml in PBS and stored in the dark at -20°C. XTT/PMS stock was prepared immediately before use by diluting the PMS 1:100 into PBS and adding 40 μ l per ml of XTT solution. Fifty microliters of XTT/PMS was added to each well of the plate and the plate was incubated for an additional 4 hours at 37°C. Adhesive plate sealers were used in place of the lids, the sealed plate was inverted several times to mix the soluble formazan product and the plate was read spectrophotometrically at A450 nm with a Molecular Devices Vmax plate reader. Using an in-house computer program % CPE (cytopathic effect) reduction, % cell viability, $IC_{25, 50 \text{ \& } 95}$, $TC_{25, 50 \text{ \& } 95}$ and other indices were calculated and the graphic results summary was displayed.

EXAMPLE 4

Reverse Transcriptase Activity Assay

5 A microtiter based reverse transcriptase (RT) reaction was utilized (Buckheit et al (1991) *AIDS Research and Human Retroviruses* 7:295-302). Tritiated thymidine triphosphate (NEN) (TTP) was resuspended in distilled H₂O at 5 Ci/ml. Poly rA and oligo dT were prepared as a stock solution
10 which was kept at -20°C. The RT reaction buffer was prepared fresh on a daily basis and consists of 125 µl 1 M EGTA, 125 µl dH₂O, 125 µl Triton X-100, 50 µl 1 M Tris (pH 7.4), 50 µl 1 M DTT, and 40 µl 1 M MgCl₂. These three solutions were mixed
15 together in a ratio of one part distilled water. Ten microliters of this reaction mixture was placed in a round bottom microtiter plate and 15 µl of virus containing supernatant was added and mixed. The plate was incubated at 37°C and
20 incubated for 60 minutes. Following reaction, the reaction volume was spotted onto filter mats, washed 6 times for 5 minutes each in a 5% sodium phosphate buffer, two times for 1 minute each in distilled water, two times for 1 minute each in
25 70% ethanol, and then dried. The dried filter mat was placed in a plastic sample bag, Betaplate scintillation fluid was added and the bag was heat-sealed. Incorporated radioactivity was quantified utilizing a Wallac Microbeta,
30 scintillation counter (Gaithersburg, MD).

EXAMPLE 5

p24 ELISA

ELISA kits were purchased from Coulter
5 (Miami, FL). The assay is performed according to
the manufacturer's recommendations. Prior to
ELISA analysis the reverse transcriptase activity
assays (described above) were routinely performed
and used the values for incorporated radioactivity
10 in the RT activity assay to determine the dilution
of our samples requires for the ELISA. Standard
curves were constructed so that the dilutions of
virus to be used in the p24 ELISA could be
accurately determined from the RT activity assay.
15 Control curves were generated in each assay to
accurately quantify the amount of capsid protein
in each sample. Data was obtained by
spectrophotometric analysis at 450 nm using a
plate reader. Molecular Devices Vmax P24
20 (Sunnydale, CA) concentrations were calculated
from the optical density values by use of the
Molecular Devices (San Jose, CA) software package
Soft Max.

EXAMPLE 6

Infectious Particles

Infectious virus particles were qualified
utilizing the CEM-SS plaque assay as described by
30 Nara et al. (*Nature* (1988) 332:469-470). Flat
bottom 96-well microtiter plates were coated with
50 μ l of poly-L-lysine (Sigma. St. Louis, MO) at
50 μ g/ml for 2 hours at 37°C. The wells were then
washed with PBS and 2.5×10^5 CEM-SS cells were

placed in the microtiter well where they became fixed to the bottom of the plate. Enough cells were added to form a monolayer of CEM-SS cells in each well. Virus containing supernatant was added from each well of the XTT plate, including virus and cell controls and each serial dilution of the test substance. The number of syncytia were qualified in the flat-bottom 96-well microtiter plate with an Olympus CK2 inverted microscope at 4 days following infection. Each syncytium resulted from a single infectious HIV virion.

EXAMPLE 7

Anti-HIV Activity in Fresh Human Cells

15

A. Assay in Fresh Human T-Lymphocytes

Fresh human peripheral blood lymphocytes (PBL) were isolated from voluntary Red Cross donors, seronegative for HIV and HBV. Leukophoresed blood was diluted 1:1 with Dulbecco's phosphate buffered saline (PBS), layered over 14 ml of Ficoll-Hypaque density gradient in a 50 ml centrifuge tube. Tubes were then centrifuged for 30 minutes at 600 X g. Banded PBLs were gently aspirated from the resulting interface and subsequently washed 2X with PBS by low speed centrifugation. After final wash, cells were enumerated by trypan blue exclusion and re-suspended at 1×10^7 /ml in RPMI 1640 with 15% Fetal Bovine Serum (FBS), 2 mM L-glutamine, 4 mg/ml PHA-P and allowed to incubate for 48 - 72 hours at 37°C. After incubation, PBLs were centrifuged and reset in RPMI 1640 with 15%

FBS, 2 mM L-glutamine, 100 U/ml penicillin, 100
μg/ml streptomycin, 10 μg/ml gentamycin, and 20
U/ml recombinant human IL-2. PBLs were maintained
in this medium at a concentration of $1-2 \times 10^6$ /ml
5 with bi-weekly medium changes, until use in assay
protocol.

For the PBL assay, PHA-P stimulated cells
from at least two normal donors were pooled, set
10 in fresh medium at 2×10^6 /ml, and plated in the
interior wells of a 96 well round bottom
microplate at 50 μL/well. Test drug dilutions
were prepared at a 2X concentration in microtiter
tubes, and 100 μl of each concentration was placed
15 in appropriate wells in a standard format. Fifty
microliters of a predetermined dilution of virus
stock was placed in each test well. Wells with
cells and virus alone were used for virus control.
Separate plates were identically set without virus
20 for drug cytotoxicity studies using an XTT assay
system.

In the standard PBL assay (MOI: 0.2), the
assay was ended on day 7 following collection of
25 cell free supernatant samples for reverse
transcriptase activity assay. In the low MOI PBL
assay (MOI: 0.02), supernatant samples were
collected on day 6, day 11, and day 14 post-
infection and analyzed for RT activity. Tritiated
30 thymidine triphosphate (NEN) (TTP) was resuspended
in distilled H₂O at 5 Ci/ml. Poly rA and oligo dT
were prepared as a stock solution which was kept
at -20°C. The RT reaction buffer was prepared
fresh on a daily basis and consists of 125 μl 1 M

DTT, and 40 μ l 1 M $MgCl_2$. These three solutions were mixed together in a ratio of 2 parts TTP, 1 part poly rA:oligo dT, and 1 part reaction buffer. Ten microliters of this reaction mixture was placed in a round bottom microtiter plate and 15 μ l of virus containing supernatant was added and mixed. The plate was incubated at 37°C in a water bath with a solid support to prevent submersion of the plate and incubated for 60 minutes. Following reaction, the reaction volume was spotted onto pieces of DE81 paper, washed 5 times for 5 minutes each in a 5% sodium phosphate buffer, 2 times for 1 minute each in distilled water, 2 times for 1 minute each in 70% ethanol, and then dried. Opti-Fluor O was added to each sample and incorporated radioactivity was quantified utilizing a liquid scintillation counter, (Wallac 1450 Microbetaplus, Gaithersburg, MD).

Tritiated thymidine incorporation was measured in parallel cultures at day 7. Each well was pulsed with 1 μ Ci of tritiated thymidine and the cells were harvested 18 hours later with a Skatron cell harvester onto glass fiber filter papers. The filters were dried, placed in a scintillation vial with 1 ml of scintillation cocktail and incorporated radioactivity was quantified on a liquid scintillation counter (e.g., Packard Tri-Carb 1900 TR450).

B. Assay in Fresh Human Monocyte-Macrophages

For isolation of adherent cells, 3×10^6 non-PHA stimulated peripheral blood cells were

resuspended in Hanks buffered saline (with calcium and magnesium) supplemented with 10% human AB serum. The cells were placed in a 24-well microtiter plate at 37°C for 2 hours. Non-
5 adherent cells were removed by vigorously washing six times. The adherent cells were cultured for 7 days in RPMI 1640 tissue culture medium with 15% fetal bovine serum. The cultures were carefully monitored for confluency during this incubation
10 period. Infection of the cells was performed with the monocytotropic HIV-1 strains BaL or ADA and the matched pair of AZT-sensitive and AZT-resistant virus isolates. Each of these virus isolates was obtained from the NIAID AIDS Research and Reference Reagent Program. High titer pools
15 of each of these viruses have been harvested from infected cultures of peripheral blood adherent cells and frozen in 1.0 ml aliquots at -80°C. Monocyte-macrophage monolayers were infected at an
20 MOI of 0.1. Compounds to be evaluated in the monocyte-macrophage assay are added to the monolayers shortly before infection in order to maximize the potential for identifying active compounds.

25
At 2 days post-infection, the medium was decanted and the cultures washed twice with complete medium in order to remove excess virus. Fresh medium alone or medium containing the
30 appropriate concentrations of drugs was added and incubation continued for an additional 5 days. XTT-tetrazolium or trypan blue exclusion assays (for cell viability) and HIV p24 ELISA assays (for production of p24 core antigen) were performed on

Day 7 post-infection. ELISA kits were purchased from Coulter. The assay is performed according to the manufacturer's recommendations. Control curves are generated in each assay to accurately quantify the amount of capsid protein in each sample. Data was obtained by spectrophotometric analysis at 450 nm using a plate reader (Molecular Devices Vmax). P24 concentrations were calculated from the optical density values by use of the Molecular Device software package Soft Max.

EXAMPLE 8

Inhibition of Acute Infection of MT-4 Cells

CPE based infection experiments were performed using MT-4 cells (Pauwels et al. (1988) *J. Virol. Meth.* 20:309; Papp et al. (1997) *AIDS Research and Human Retroviruses In Press*). MT-4 cells were obtained from the AIDS Research and Reference Reagent Bank, Division of AIDS, NIAID, NIH contributed Dr. Richman (Pauwels et al. (1988) *J. Virol. Meth.* 20:309). T-lymphoid H9 (HUT-78) cells were obtained from Dr. Robert Gallo, National Cancer Institute, Bethesda, MD (Popovic et al. (1984) *Science* 224:497; Gazdar et al. (1980) *Blood* 55:409). Cell cultures were maintained in RPMI 1640 medium (GIBCO Laboratories, Grand Island, NY) supplemented with 20% (H9 cells), or 10% (MT-4 cells) heat-inactivated fetal bovine serum (Sigma Chemical Co., St. Louis, MO) 250 units/ml penicillin, 250 µg/ml streptomycin, 2 mM l-glutamine, and 10 mM HEPES(N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid) buffer (complete medium) at

37°C in 5% CO₂. HIV-1 IIB was originally obtained from Dr. Robert Gallo, National Cancer Institute (Popovic (1984) *Science* 224:497). Virus stocks of HIV-1 were prepared from cell-free filtered supernatant of chronically infected H9 cultures by the shaking method as previously described by Vujoie et al. (*J. Infectious Diseases* (1988) 157:1047).

Experiments were performed under two sets of conditions. Dilutions of hybrid oligonucleotides according to the invention having SEQ ID NO:1 were prepared in 96-well plates and infections were performed either in the presence of inhibitors, by adding MT-4 cells and a TCID_{CPE-90%} concentration of HIV_{-HHB} directly to the wells, or by infecting MT-4 cells for 4 hours at 37°C in the absence of inhibitors, washing to remove non-adsorbed virus, then adding the infected cells to wells containing inhibitors. The cultures were incubated for 6 days and CPE measured using the MTT dye method. (Rapid, (1983) *J. Immunolog. Meth.* 65:55).

The results demonstrate that an oligonucleotide of the invention inhibits HIV-1 infection when added to cells during viral infection (FIG. 1) or post-viral adsorption (FIG. 2).

EXAMPLE 9

Measurement of Orally Administered Oligonucleotide

A. Animals and Treatment

Male Sprague-Dawley rats (100-120 g, Harlan Laboratories, Indianapolis, IN) and male CD-/F2 mice (25 ± 3 g, Charles River Laboratory, Wilmington, MA) are used in the study. The
5 animals are fed with commercial diet and water *ad libitum* for 1 week prior to the study.

Unlabelled and ³⁵S-labelled oligonucleotides are dissolved in physiological saline (0.9% NaCl) in a concentration of 25 mg/ml, and are
10 administered to the fasted animals via gavage at the designated dose (30-50 mg/kg for rats and 10 mg/kg for mice). Doses are based on the pretreatment body weight and rounded to the
15 nearest 0.01 ml. After dosing, each animal is placed in a metabolism cage and fed with commercial diet and water *ad libitum*. Total voided urine is collected and each metabolism cage is then washed following the collection intervals.
20 Total excreted feces is collected from each animal at various timepoints, and feces samples are homogenized prior to quantitation of radioactivity. Blood samples are collected in heparinized tubes from animals at the various
25 timepoints. Plasma is separated by centrifugation. Animals are euthanized by exsanguination under sodium pentobarbital anesthesia at various times (i.e., 1, 3, 6, 12, 24, and 48 hr; 3 animals/time point). Following
30 euthanasia, the tissues are collected from each animal. All tissues/organs are trimmed of extraneous fat or connective tissue, emptied and cleaned of all contents, individually weighed, and the weights recorded.

To quantitate the total absorption of the hybrid oligonucleotide, two additional groups of animals (3/group) for each test oligonucleotide are treated using the same procedure as above.

5 Animals are killed at 6 or 12 hr post dosing, and the gastrointestinal tract is then removed. Radioactivities in the gastrointestinal tract, feces, urine, plasma, and the remainder of the body is determined separately. Total recovery of
10 radioactivity is also determined to be $95 \pm 6\%$. The percentage of the absorbed hybrid oligonucleotide-derived radioactivity is determined by the following calculation:
15
$$\frac{(\text{total radioactivity in the remainder of the body} + \text{total radioactivity in urine})}{(\text{total radioactivity in the gastrointestinal tract, feces, urine, plasma, and the remainder of the body})}$$

20 B. Radioactive Labelling of Oligonucleotide

To obtain ^{35}S -labelled oligonucleotide, synthesis is carried out in two steps. The first nucleotides of the oligonucleotide sequence from
25 its 3'-end are assembled using the β -cyanoethyl-phosphoramidite approach (see, Beaucage in *Protocols for Oligonucleotides and Analogs* (Agrawal, ed.), Humana Press, (1993), pp. 33-61). The last nucleotides are assembled using the H-phosphonate approach
30 (see, Froehler in *Protocols for Oligonucleotides and Analogs* (Agrawal, ed.) Humana Press, 1993, pp. 63-80). Controlled pore glass (CPG) support-bound oligonucleotide (30 mg of CPG; approximately $1 \mu\text{M}$) containing five H-phosphonate linkage is oxidized

with $^{35}\text{S}_8$ (4 mCi, 1 Ci/mg, Amersham; 1 Ci = 37 GBq) in 60 ml carbon disulfide/pyridine/triethylamine (10:10:1). The oxidation reaction is performed at room temperature for 1 hr with occasional shaking. Then 2 μl , 5 μl , and 200 μl of 5% cold sulfur ($^{32}\text{S}_8$) in same solvent mixture is added every 30 min to complete the oxidation. The solution is removed and the CPG support is washed with carbon disulfide/pyridine/ triethylamine (10:10:1) (3 x 500 μl) and with acetonitrile (3 x 700 μl). The product is deprotected in concentrated ammonium hydroxide (55°C, 14 hr) and evaporated. The resultant product is purified by polyacrylamide gel electrophoresis (20% polyacrylamide containing 7 M urea). The desired band is excised under UV shadowing and the PS-oligonucleotide was extracted from the gel and desalted with a Sep-Pak C18 cartridge (Waters) and Sephadex G-15 column.

C. Total Radioactivity Measurements

The total radioactivities in tissues and body fluids is determined by liquid scintillation spectrometry (LS 6000TA, Beckman, Irvine, CA). In brief, biological fluids (plasma, 50-100 μl ; urine, 50-100 μl) are mixed with 6 ml scintillation solvent (Budget-Solve, RPI, Mt. Prospect, IL) to determine total radioactivity. Feces are ground and weighed prior to being homogenized in a 9-fold volume of 0.9% NaCl saline. An aliquot of the homogenate (100 μl) is mixed with solubilizer (TS-2, RPI, Mt. Prospect, IL) and then with scintillation solvent (6 ml) to permit quantitation of total radioactivity.

Following their removal, tissues are immediately blotted on Whatman No. 1 filter paper and weighed prior to being homogenized in 0.9% NaCl saline (3-5 ml per gram of wet weight). The resulting homogenate (100 μ l) is mixed with solubilizer (TS-2, RPI, Mt. Prospect, IL) and then with scintillation solvent (6 ml) to determine total radioactivity. The volume of 0.9% NaCl saline added to each tissue sample is recorded. The homogenized tissues/organs are kept frozen at $\leq -70^{\circ}\text{C}$ until the use for further analysis.

D. HPLC Analysis

The radioactivity in urine is analyzed by paired-ion HPLC using a modification of the method described essentially by Sands et al. (*Mol. Pharm.* (1994) 45:932-943). Urine samples are centrifuged and passed through a 0.2- μm Acro filter (Gelman, Ann Arbor, MI) prior to analysis. Hybrid oligonucleotide and metabolites in plasma samples are extracted using the above methods in sample preparation for PAGE. A Microsorb MV-C4 column (Rainin Instruments, Woburn, MA) is employed in HPLC using a Hewlett Packard 1050 HPLC with a quaternary pump for gradient making. Mobile phase includes two buffers; Buffer A was 5 mM-A reagent (Waters Co., Bedford, MA) in water and Buffer B is 4:1 (v/v) Acetonitrile (Fisher)/water. The column is eluted at a flow rate of 1.5 ml/min, using the following gradient: (1) 0-4 min, 0% buffer B; (2) 4-15 min 0-35% Buffer B; and (3) 15-70 min 35%-80% Buffer B. The column is equilibrated with Buffer A for at least 30 min prior to the next run. By

5 using a RediFrac fraction collector (Pharmacia LKB Biotechnology, Piscataway, NJ), 1-min fractions (1.5 ml) are collected into 7-ml scintillation vials and mixed with 5 ml scintillation solvent to determine radioactivity in each fraction.

E. Analysis of Test Oligonucleotides

10 Polyacrylamide gel electrophoresis (PAGE) of oligonucleotides and its metabolites is carried out by known and established methods. Plasma and tissue homogenates are incubated with proteinase K (2 mg/ml) in extraction buffer (0.5% SDS/10 mM NaCl/20 mM Tris-HCl, pH 7.6/10 mM EDTA) for 1 hr
15 at 60°C. The samples are then extracted twice with phenol/chloroform (1:1, v/v) and once with chloroform. After ethanol precipitation, the extracts are analyzed by electrophoresis in 20% polyacrylamide gels containing 7 M urea. Urine
20 samples are filtered, desalted and then analyzed by PAGE. The gels are fixed in 10% acetic acid/10% methanol solution and then dried before autoradiography.

25 EQUIVALENTS

Those skilled in the art will recognize, or be able to ascertain, using no more than routine experimentation, numerous equivalents to the
30 specific substances and procedures described herein. Such equivalents are considered to be within the scope of this invention, and are covered by the following claims.